

# SABOURAUD DEXTROSE AGAR WITH GENTAMYCIN+CHLORAMPHENICOL

**INTENDED USE:**

SABOURAUD DEXTROSE AGAR + CHLORAMPHENICOL + GENTAMICIN is a selective medium that can be used for the cultivation of yeast, molds and aciduric microorganisms.

**PRINCIPLE AND INTERPRETATION:**

This medium is a modification enriched with Gentamicin and Chloramphenicol which supports the growth of a wide range of fungi and, due to its antibiotic content, inhibits the great majority of bacterial contaminants. Dextrose contained in Saboraud dextrose is the fermentable carbohydrate providing carbon and energy. Peptone mixture provides nitrogen, vitamins, minerals and amino acids essential for growth. Bacteriological agar is the solidifying agent. The high dextrose concentration and acidic pH make this medium selective for fungi. Chloramphenicol is an antibiotic which aids in isolating pathogenic fungi from heavily contaminated material, as it inhibits most contaminating bacteria. It is a recommended antibiotic for use with media due to its heat stability and wide bacterial spectrum. Gentamicin is an aminoglycoside antibiotic, used to treat many types of bacterial infections by inhibiting its growth.

**COMPOSITION:**

Ingredients	Gr/Liter
Pancreatic Digest of Casein	5 gr
Peptic Digest of Animal Tissue	5 gr
Dextrose	40 gr
Chloramphenicol	0,05 gr
Gentamicin.	0,05 gr
Agar	15 gr

\*\*\*Formula adjusted, standardized to suit performance parameters

pH: 5,6 ± 0,2

**PRECAUTIONS:**

For professional use only. Do not use plates if they show evidence of microbial contamination, discoloration, drying, cracking or other signs of deterioration.

**TEST PROCEDURE:**

Allow the plates to warm to room temperature and the agar surface to dry before inoculating. Heavily inoculate and streak the specimen as soon as possible after collection. If the specimen to be cultured is on a swab, roll the swab over a small area of the agar surface. Streak for isolation with a sterile loop. Examine colonial morphology.

Inoculate two samples for isolation of fungi that cause systemic mycoses; incubate one at 25-30°C and the other set at 35± 2°C. Examine cultures weekly for a period of up to four to six weeks; solid media should be incubated under conditions of increased humidity during prolonged incubation.

**QUALITY CONTROL:**
**1.Sterility Control:**

Incubation 72 hours at 20-25°C and 48 hours at 30-35°C: NO GROWTH

**2.Physical/Chemical Control**

pH: 5,6 ± 0,2

Appearance: Light amber

**3.Microbiological Control: Incubation at 25±2 °C during:48 h -5 d**

Microorganism	Inoculum (CFU)	Results
		Growth
Candida albicans ATCC 10231	10-100	Good
Aspergillus niger ATCC 16404	10-100	Good
Escherichia coli ATCC 25922	100-1000	Inhibition
Staphylococcus aureus ATCC 25923	100-1000	Inhibition

## LIMITATIONS OF THE PROCEDURE:

Some fungi may be inhibited by the antibiotics in these media. For identification, organisms must be in pure culture. Morphological, biochemical, and/or serological tests should be performed for final identification. Consult appropriate texts for detailed information and recommended procedures. A single medium is rarely adequate for detecting all organisms of potential significance in a specimen. It should be recognized that organisms generally susceptible to the antimicrobial agent in a selective medium may be completely or only partially inhibited depending upon the concentration of the agent, the characteristics of the microbial strain and the number of organisms in the inoculum. Organisms that are generally resistant to the antimicrobial agent should not be inhibited. Cultures of specimens grown on selective media should, therefore, be compared with specimens cultured on nonselective media to obtain additional information and help ensure recovery of potential pathogens.

## STORAGE CONDITIONS AND SHELF LIFE:

Store the prepared medium at 2 - 12°C. Use before expiry date on the label..Do not use beyond stated expiry date.

## DISPOSAL:

Incubated prepared medium may contain active bacteria and micro-organisms. Do not open infected medium. Infected plate should be autoclaved, incinerated or opened and soaked in a chlorine-based disinfectant (liquid bleach) for 20 minutes prior to disposal.

## PACKAGING:

**Katalog Number:** 02043

**Packaging:** Single wrap

**Content:** 10 plates/each package

## REFERENCES:

1. Sabouraud, R. 1892. Contribution a l'etude de la trichophytie humaine. Etude clinique, microscopique et bacteriologique sur la pluralite des trichophyton de l'homme. Ann. Dermatol. Syphil. 3:1061-1087.
2. Haley, L.D., J. Trandel, and M.B. Coyle. 1980. Cumitech 11, Practical methods for culture and identification in the clinical microbiology laboratory. Coordinating ed., J.C. Sherris. American Society for Microbiology, Washington, D.C.
3. Ajello, L., L.K. Georg, W. Kaplan, and L. Kaufman. 1963. CDC laboratory manual for medical mycology. PHS Publication No. 994, U.S. Government Printing Office, Washington, D.C.
4. Kane, J., and R.C. Summerbell. 1999. Trichophyton, Microsporum, Epidermophyton, and agents of superficial mycoses, . 1275-1294. In P.R. Murray, E.J. Baron, M.A. Pfaller, F.C. Tenover, and R.H. Tenover (ed.), Manual of clinical microbiology, 7th ed. American Society for Microbiology, Washington, D.C.
5. Lorian, V. (ed.). 1996. Antibiotics in laboratory medicine, 4th ed. Williams & Wilkins, Baltimore.
6. Clinical and Laboratory Standards Institute. 2005. Approved Guideline M29-A3. Protection of laboratory workers from occupationally acquired infections, 3rd ed. CLSI, Wayne, PA.



Aseptic Sterile



Batch Code



Catalogue Number



Negative Controls



Positive Controls



Use by



Temperature  
Limitation



Do not reuse



Contains sufficient  
for <n> tests



Look at user manual



Manufacturer