

HYGİSLİDE MALT EXTRACT AGAR / SDA

PRINCIPLE AND INTERPRETATION:

Side1: MEA: Malt Extract Agar is recommended for the detection, isolation and enumeration of fungi. Malt extract contains polysaccharides which are used as energy source. It makes the medium acidic as well. Mycological peptone serves as a nitrogen source.

Side2: SDA: An acidic pH medium for the isolation of dermatophytes, other fungi and yeasts. Sabouraud Dextrose Agar is a peptone medium supplemented with dextrose to support the growth of fungi. The peptones are sources of nitrogenous growth factors. Dextrose provides an energy source for the growth of microorganisms.

COMPOSITION:

MEA

Ingredients	Gr/Liter
Malt extract	30 gr
Mycological peptone	5 gr
Agar	9 gr

pH: 5,4 ± 0,2

SDA

Ingredients	Gr/Liter
Mycological peptone	10 gr
Glucose(dextrose)	40 gr
Agar	15 gr

pH: 5,6 ± 0,2

***Formula adjusted, standardized to suit performance parameters

INSTRUCTIONS FOR USE:

Testing Fluids:

1. Mix liquid test sample.
2. Remove the paddle from the vial. Do not touch the agar surfaces.
3. Immerse the slide in the fluid to be tested for about 5- 10 seconds. Alternatively expose the slide to a spray or running fluid so that the slide surfaces are covered.
4. Both agar surfaces must be completely contacted.
5. Allow excess fluid to drain off both paddle agar surfaces.
6. Replace the Slide into the tube and twist to tighten the cap. Label the tube with the identification label supplied. Incubate the slide as directed later.

Testing Surfaces:

1. Remove the paddle from the vial. Do not touch the agar surfaces.
2. To assure an accurate area recovery, contact the paddle to 20²cm of the surface by contacting the surface twice in separate 10²cm areas.
3. Replace the Slide into the tube and twist to tighten the cap. Label the tube with the identification label supplied. Incubate the slide as directed later.

QUALITY CONTROL:

1.Sterility Control:

Incubation 2 d at 30-35°C and 3 d at 20-25°C: NO GROWTH

2.Physical/Chemical Control

	pH	Apperance:
MEA:	5,4 ± 0,2	Light amber
SDA:	5,6 ± 0,2	Amber

3.Microbiological Control: Incubate at 35±2 °C temperature for 24 hours and 25±2 °C 48 h-5 days.

Side1: MEA

Microorganism	Inoculum (CFU)	Results	
		Growth	Reaction
Candida albicans ATCC 10231	10-100	Good	Good
Aspergillus niger ATCC 16404	10-100	Good	Good

Side2: SDA

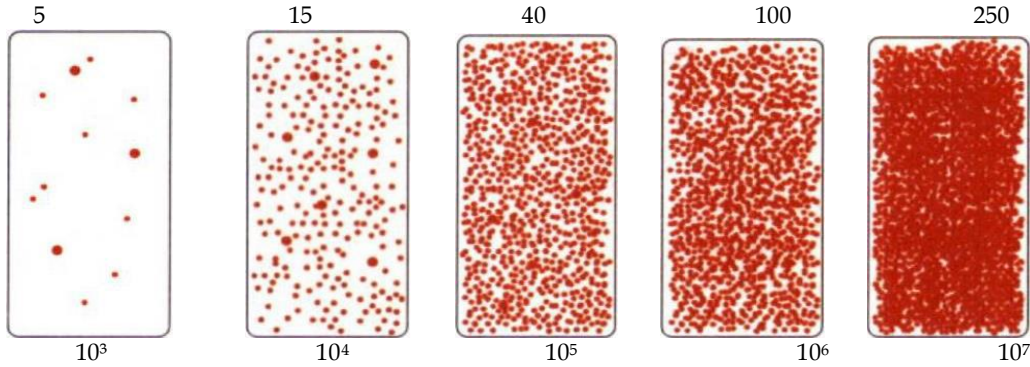
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Aspegillus brasiliensis ATCC 16404	10-100	Good	Good

INTERPRETATION OF RESULTS

Compare the slide surfaces against the comparison chart printed below. Read the result corresponding to fluids or surfaces as appropriate. Note that very high levels of organisms could lead to a confluent growth and could be recorded as a nil result. Compare against an unused slide when reading results.

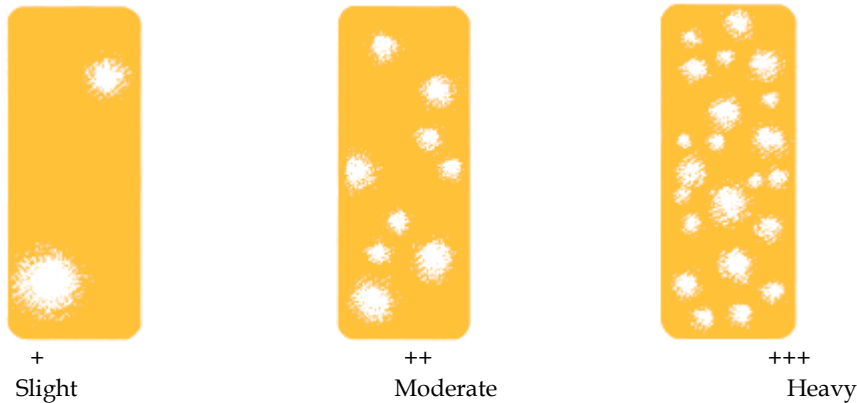
Bacteria Comparison Chart

Surfaces
CFU/cm²



Fluids
CFU/mL

Fungi Comparison Chart



DISPOSAL:

Incubated Slides may contain active bacteria and micro-organisms. Do not open infected slides except as part of disposal procedure. Infected slides should be autoclaved, incinerated or opened and soaked in a chlorine-based disinfectant (liquid bleach) for 20 minutes prior to disposal.

STORAGE CONDITIONS AND SHELF LIFE:

Slides should be stored in 2-20 °C. Temperature fluctuations may result in condensation settling at the bottom of the vial, although this does not affect culture properties, it could reduce the shelf-life or cause the agar to separate from the plastic paddle support.

Avoid sudden temperature changes. Shield from direct sunlight. Do not allow paddles to freeze. Do not use any slides which have been inadvertently contaminated during storage and which are already showing growth of micro-organisms

Use before expiry date on the label. Do not use beyond stated expiry date.

PACKAGING:

Katalog Number: 06006

Content/Packaging: 20 Slides/Box

REFERENCES:

1. American Public Health Association, Standard Methods for the Examination of Dairy Products, 14th ed., APHA Inc., Washington, D.C. (1978)
2. E.W. Frampton, et al., Comparison of β -glucuronidase and indole-based direct plating methods for enumeration of unstressed E. coli, J. Food Protect. 53, 933 (1990)
3. Carlier Gwendoline I. M. (1948) Brit. J. Derm. Syph. 60. 61-63.
4. Hodges R. S. (1928) Arch. Derm. Syph., New York, 18. 852.
5. Sabouraud R. (1910) 'Les Teignes', Masson, Paris.
6. Georg Lucille K., Ajello L. and Papageorge Calomira (1954) J. Lab. Clin. Med. 44. 422-428.
7. Ajello Libero (1957) J. Chron. Dis. 5. 545-551.
8. Williams Smith H. and Jones J. E. T. (1963) J. Path. Bact. 86. 387-412.
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Aseptic Sterile



Batch Code



Catalogue Number



Negative Controls



Positive Controls



Use by



Temperature Limitation



Do not reuse



Contains sufficient for <n> tests



Look at user manual



Manufacturer