

SURCHECK TRYPTIC SOY AGAR W/ 4 NEUTRALISANT

INTENDED USE:

General purpose medium for isolation and culture of microorganisms with neutralisers.

PRINCIPLE AND INTERPRETATION:

The unsupplemented medium is not used as a primary isolation medium for clinical applications.

In Tryptic Soy Agar, the combination of casein and soy peptones renders the medium nutritious by supplying organic nitrogen, particularly amino acids and longer-chained peptides. Sodium chloride maintains the osmotic equilibrium. Agar is the solidifying agent.

The neutralizing additives Polysorbate (Tween) 80, glycine, sodium Thiosulfate, histidine, lecithin and sodium thioglycollate reverse the growth inhibitory effect of most disinfectants and antiseptics. Neutralizing efficiency of histidine could be shown against formaldehyde and formaldehyde releasing agents. Na-thiosulfate inactivates sodium hypochlorite and acidified sodium chlorite. Lecithin, particularly in combination with Polysorbate 80, is effective against quaternary ammonium compounds, amphoteric surfactants, benzamidines, chlorhexidines and dequadin. Polysorbate 80 inactivates benzyl alcohol, carbanilides, dichlorbenzyl alcohols, benzoic acid, p-hydroxybenzoic acid and its esters, phenols, phenylethyl alcohols and Solbrols.

COMPOSITION:

Ingredients	Gr/Liter
Pancreatic digest of casein	15 gr
Enzymatic* digest of soya bean	5 gr
Sodium chloride	5 gr
Tween 80 (polysorbate 80)	5 gr
Lecithin	0,7 gr
Histidine	1 gr
Sodium Thiosulfate	0,5 gr
Agar	15 gr

***Formula adjusted, standardized to suit performance parameters

pH: 7,3 ± 0.2

PRECAUTIONS:

For professional use only. Do not use plates if they show evidence of microbial contamination, discoloration, drying, cracking or other signs of deterioration.

TEST PROCEDURE:

Before use, agar surface should be smooth and moist, but without excessive moisture because this could be cause of confluent growth.

1. The medium should be warmed to room temperature and the surface should be dry prior to use.
2. Open the lid carefully.
3. Press the convex agar for 10 seconds on the surface to be tested with a light uniform pressure. Pressure should be the same for every sample. Do not move plate laterally.
4. Replace the lid and mark the plate with appropriate data.
5. Clean the sample area with a disinfectant in order to any remaining of the agar.
6. The plates for determination of the total aerobic bacterial count should be incubated at 30 to 35 °C for 48 to 72 hours, while the plates for determination of the total yeast and mold count should be incubated at 20 to 25 °C for 5 to 7 days

QUALITY CONTROL:**1.Sterility Control:**

Incubation 2d at 30-35°C and 3d at 20-25°C: NO GROWTH

2.Physical/Chemical Control

pH: 7,3 ± 0,2

Apperance: Light amber

3. Microbiological Control: Incubation at 35± 2 °C;24-48 hours and 25±2 °C:5 d

Microorganism	Inoculum (CFU)	Results	
		Growth	Reaction
Bacillus subtilis ATCC 6633	10-100	Good	>70 %
Staphylococcus aureus ATCC 6538	10-100	Good	>70 %
Candida albicans ATCC 10231	10-100	Good	>70 %
Pseudomonas aeruginosa ATCC 9027	10-100	Good	>70 %
Aspergillus brasiliensis ATCC 16404	10-100	Good	>70 %
Escherichia coli ATCC 8739	10-100	Good	>70 %
Staphylococcus epidermidis ATCC 12228	10-100	Good	>70 %

LIMITATIONS OF THE PROCEDURE:

Due to nutritional variation, some strains may be encountered that grow poorly or fail to grow on this medium.

STORAGE CONDITIONS AND SHELF LIFE:

Store the prepared medium at 2-12°C or 2-25°C. Use before expiry date on the label. Do not use beyond stated expiry date.

DISPOSAL:

Incubated prepared medium may contain active bacteria and micro-organisms. Do not open infected medium. Infected plate should be autoclaved, incinerated or opened and soaked in a chlorine-based disinfectant (liquid bleach) for 20 minutes prior to disposal.

PACKAGING:

Katalog Number: 06127

Packaging: Single wrap

Content: 10 plates/each package

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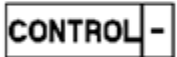
Aseptic Sterile



Batch Code



Catalogue Number



Negative Controls



Positive Controls



Use by



Temperature Limitation



Do not reuse



Contains sufficient for <n> tests



Look at user manual



Manufacturer