

HYGİSLİDE SDA / CHROMAGAR ECC

PRINCIPLE AND INTERPRETATION:

Side1: SDA: An acidic pH medium for the isolation of dermatophytes, other fungi and yeasts. Sabouraud Dextrose Agar is a peptone medium supplemented with dextrose to support the growth of fungi. The peptones are sources of nitrogenous growth factors. Dextrose provides an energy source for the growth of microorganisms.

Side2: Chromagar ECC: Chromogenic medium for the detection and enumeration of β -glucuronidase positive E.coli and coliforms in food and water samples. Coliforms, Enterobacteriaceae able to ferment lactose (lactose positive Enterobacteriaceae), are bacteria present in human and warm blooded animals intestinal flora, in the soil and water. Coliforms are proof of organic, environmental or faecal contamination. Faecal contamination, due to coliforms coming from animal waste, consists mainly of Escherichia coli and thermotolerant Klebsiella. Strict regulations exist for E.coli/Coliform presence in water and food samples. This can be explained by the importance of these germs in determining water and food safety.

COMPOSITION:

SDA

Ingredients	Gr/Liter
Mycological peptone	10 gr
Glucose(dextrose)	40 gr
Agar	15 gr

pH: 5,6 \pm 0,2

Chromagar ECC

Ingredients	Gr/Liter
Peptone and yeast extract	8 gr
NaCl	5 gr
Chromogenic mix	4,8 gr
Agar	15 gr

pH: 7,2 \pm 0,2

***Formula adjusted, standardized to suit performance parameters

INSTRUCTIONS FOR USE:

Testing Fluids:

1. Mix liquid test sample.
2. Remove the paddle from the vial. Do not touch the agar surfaces.
3. Immerse the slide in the fluid to be tested for about 5- 10 seconds. Alternatively expose the slide to a spray or running fluid so that the slide surfaces are covered.
4. Both agar surfaces must be completely contacted.
5. Allow excess fluid to drain off both paddle agar surfaces.
6. Replace the Slide into the tube and twist to tighten the cap. Label the tube with the identification label supplied. Incubate the slide as directed later.

Testing Surfaces:

1. Remove the paddle from the vial. Do not touch the agar surfaces.
2. To assure an accurate area recovery, contact the paddle to 20²cm of the surface by contacting the surface twice in separate 10²cm areas.
3. Replace the Slide into the tube and twist to tighten the cap. Label the tube with the identification label supplied. Incubate the slide as directed later.

QUALITY CONTROL:

1.Sterility Control:

Incubation 2 d at 30-35°C and 3 d at 20-25°C: NO GROWTH

2.Physical/Chemical Control

	pH	Apperance:
SDA:	5,6 \pm 0,2	Amber
Chromagar ECC:	7,2 \pm 0,2	Amber

3.Microbiological Control: Incubate 48 h-5 days at 20-25 °C and 24-48 hours at 35±2 °C.

Side1: SDA

Microorganism	Inoculum (CFU)	Results	
		Growth	Reaction
Candida albicans ATCC 10231	10-100	Good	Good
Aspegillus brasiliensis ATCC 16404	10-100	Good	Good

Side2: Chromagar ECC

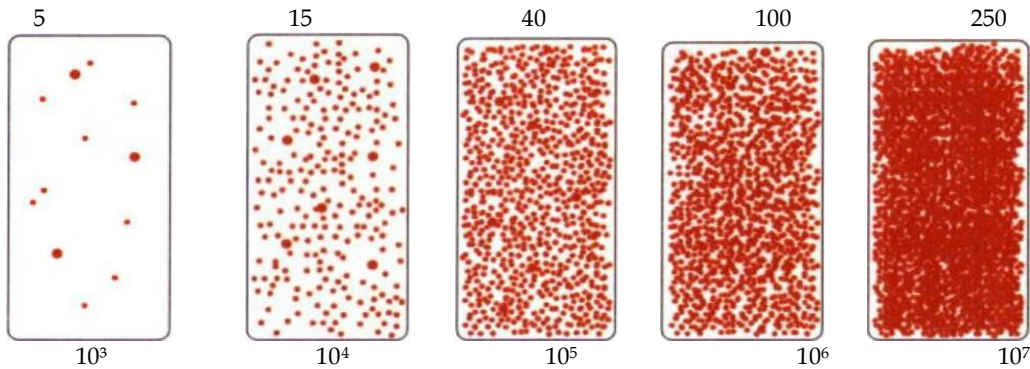
Microorganism	Inoculum (CFU)	Results	
		Growth	Reaction
E.coli ATCC 25922	10-100	Growth	Blue
Citrobacter freundii ATCC 8090	10-100	Growth	Mauve
E.cloacae ATCC 43560	10-100	Growth	Mauve
E.aerogenes ATCC 13048	10-100	Growth	Mauve
K.pneumoniae ATCC 4352	10-100	Growth	Mauve
Staphylococcus aureus ATCC 25923	100-1000	Inhibition	-
Enterococcus faecalis ATCC 25212	100-1000	Inhibition	-

INTERPRETATION OF RESULTS

Compare the slide surfaces against the comparison chart printed below. Read the result corresponding to fluids or surfaces as appropriate. Note that very high levels of organisms could lead to a confluent growth and could be recorded as a nil result. Compare against an unused slide when reading results.

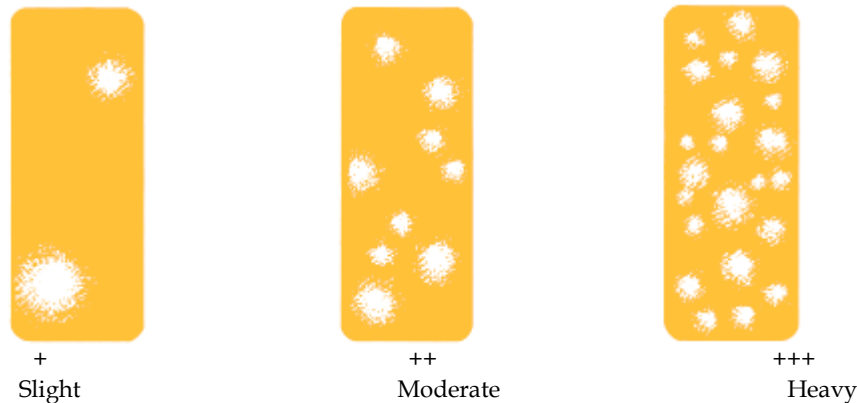
Bacteria Comparison Chart

Surfaces
CFU/cm²



Fluids
CFU/mL

Fungi Comparison Chart



DISPOSAL:

Incubated Slides may contain active bacteria and micro-organisms. Do not open infected slides except as part of disposal procedure. Infected slides should be autoclaved, incinerated or opened and soaked in a chlorine-based disinfectant (liquid bleach) for 20 minutes prior to disposal.

STORAGE CONDITIONS AND SHELF LIFE:

Slides should be stored in 2-20 °C. Temperature fluctuations may result in condensation settling at the bottom of the vial, although this does not affect culture properties, it could reduce the shelf-life or cause the agar to separate from the plastic paddle support.

Avoid sudden temperature changes. Shield from direct sunlight. Do not allow paddles to freeze. Do not use any slides which have been inadvertently contaminated during storage and which are already showing growth of micro-organisms

Use before expiry date on the label. Do not use beyond stated expiry date.

PACKAGING:

Katalog Number: 06028

Content/Packaging: 20 Slides/Box

REFERENCES:

1. Carlier Gwendoline I. M. (1948) Brit. J. Derm. Syph. 60. 61-63.
2. Hodges R. S. (1928) Arch. Derm. Syph., New York, 18. 852.
3. Sabouraud R. (1910) `Les Teignes', Masson, Paris.
4. Georg Lucille K., Ajello L. and Papageorge Calomira (1954) J. Lab. Clin. Med. 44. 422-428.
5. Ajello Libero (1957) J. Chron. Dis. 5. 545-551.
6. Williams Smith H. and Jones J. E. T. (1963) J. Path. Bact. 86. 387-412.
7. Hantschke D. (1968) Mykosen. 11. 113-115
8. Baird-Parker A. C. (1962) J. Appl. Bact. 25. 12-19.
9. Zebovitz E., Evans J. B. and Niven C. F. (1955) J. Bact. 70. 686-689.
10. Baird-Parker A. C. (1963) J. Gen. Microbiol. 30. 409-413. Shaw S., Scott M. and Cowan T. (1957) J. Gen. Microbiol. 5. 1010-1023.
11. 2005 Submitted to: International Association for Food Protection Publication Date: July 15, 2005 Citation: Bailey, J.S., Cray, P.J., Berrang, M.E., Plumblee, J. 2005. Comparison of petrifilm and chromagar ecc for isolation of e. coli from chicken [abstract]. International Association for
12. 2001 2001, National Institute of Industrial Technology
13. 1999 Alonso J. L. et al 1999. Applied and Environmental Microbiology, 65: 3746-3749.
14. A comparative study of selective media used to detect and confirm coliforms and Escherichia coli in water samples using membrane filtration 1995 1995. Abstract by Collyer J.



Aseptic Sterile



Batch Code



Catalogue Number



Negative Controls



Positive Controls



Use by



Temperature Limitation



Do not reuse



Contains sufficient for <n> tests



Look at user manual



Manufacturer